

***In vitro* reactivation of trichlorfon-inhibited butyrylcholinesterase using HI-6, obidoxime, pralidoxime and K048**

MIROSLAV POHANKA^{1,2,3}, DANIEL JUN^{1,3} & KAMIL KUČA^{1,3}

¹Faculty of Military Health Sciences, Center of Advanced Studies, University of Defence, Hradec Kralove, Czech Republic, ²Center of Biological Defence, Central Military Institute of Health, Techonin, Czech Republic, and ³Department of Toxicology, Faculty of Military Health Sciences, University of Defense, Hradec Kralove, Czech Republic

(Received 11 March 2008; accepted 19 May 2008)

Abstract

Trichlorfon is a specific inhibitor of cholinesterases. It was typically used as an insecticide; however, trichlorfon was described as useful for symptomatic treatment of Alzheimer's disease some years ago. The presented study is aimed at reactivation of trichlorfon-inhibited butyrylcholinesterase since this enzyme play an important role in Alzheimer's disease as deputy for acetylcholinesterase and furthermore it could be applied as a scavenger in case of overdosing. We used *in vitro* reactivation test for considering only reactivation efficacy of butyrylcholinesterase that is inhibited by trichlorfon and not reactivation of butyrylcholinesterase inhibited by trichlorfon metabolic products. Four reactivators were used: HI-6, pralidoxime, obidoxime, and K048. Although all of the reactivators seem to be effective at 1 mM concentration, a lower concentration was not able ensure sufficient reactivation. There was also an observed lowering of reactivation efficacy when butyrylcholinesterase was exposed to trichlorfon for a longer time interval.

Keywords: *metrifonate, chlorophos, Alzheimer's disease, reactivation, oxime, butyrylcholinesterase, trichlorfon*

Introduction

Trichlorfon is an organophosphonate pesticide commercialized under lots of trade names including the well known metrifonate and chlorophos. It is an inhibitor of important enzyme in neurosynapses: acetylcholinesterase (AChE; EC 3.1.1.7). According to the IUPAC, the correct name of this compound is following: (RS)-2,2,2-trichloro-1-(dimethoxyphosphinoyl)ethanol. Common application of trichlorfon is treatment of vegetable, fruit, and forest and so on when damaging by insect occurs. Trichlorfon is also well known medicament for effective suppression of cognitive and behavioral symptoms of Alzheimer's disease [1–3]. The efficacy and low side effects of trichlorfon were proved on a total of 408 patients [4]; on the other side, trichlorfon could induce aneuploidy in male mouse germ cells [5]. After intake by organism, trichlorfon is spontaneously transformed

into 2,2- dimethyl dichlorovinyl phosphate that is more affine to serine in AChE active centre than the original trichlorfon [6,7].

Reactivation of cholinesterases inhibited by organophosphates and organophosphonates is needed when intoxication occurs; reactivators used for reactivation purposes are compounds with oxime functional group able to dissociate phosphate or phosphonate from cholinesterase [8]. Because of evolutionary rigid structure of cholinesterases, the similar reactivation efficacy is widely expected for cholinesterases from various organisms [9]. Wide group of reactivators is commonly known, especially obidoxime, pralidoxime, and HI-6 are the commercialized ones; however, new reactivators are referred, too [10–16].

The presented study follows the efficacy of butyrylcholinesterase (BChE; EC 3.1.1.8) reactivation using currently available reactivators. BChE plays

Correspondence: K. Kuca, Faculty of Military Health Sciences, Center of Advanced Studies, University of Defence, Hradec Kralove, Czech Republic. Tel: 420973251523. Fax: 420495518094. E-mail: kucakam@pmfhk.cz

substitute role in organism [17], especially when AChE is inhibited by low dose of trichlorfon for treatment purposes. Previously described photometric method was chosen as a convenient tool for reactivation monitoring [18]. The *in vitro* method was chosen for possibility of monitoring reactivation of only trichlorfon and not its metabolic products since in some cases of acute intoxication such as overdosing in Alzheimer's disease treatment could be trichlorfon harmful itself.

Material and methods

Chemicals

Trichlorfon in analytical purity was purchased from Labor Dr. Ehrenstorfer-Schafers (Augsburg, Germany). Acetylthiocholine chloride (ATChCl), 5,5'-dithiobis (2-nitrobenzoic acid) (DTBN), and BChE from human serum were purchased from Sigma-Aldrich (Czech Republic branch). Pralidoxime (1-methyl-2-hydroxyiminomethylpyridinium chloride), obidoxime (1,3-bis(4-hydroxyiminomethylpyridinium)-2-oxapropane dichloride), HI-6 (1-(2-hydroxyiminomethylpyridinium)-3-(4-carbamoylpyridinium)-2-oxapropane dichloride) and K048 (1-(4-hydroxyiminomethylpyridinium)-4-(4-carbamoylpyridinium)-butane dibromide) were previously synthesized accordingly to references [19–22]. Used reactivators are presented in Figure 1. All substances were diluted by phosphate buffered saline (PBS).

Measuring format

Plates with 96-wells (Gama, Ceske Budejovice, Czech Republic) were used throughout all experiments. BChE was adjusted up 10 nkat/mL by PBS dilution. 10 μ L of BChE solution and 10 μ L of trichlorfon (or 10 μ L of PBS as negative control) solution (0.5 mM)

were mixed together within one well and let incubate for 0.5 h or for 1 h. Then 1 mM ATChCl with DTBN 0.4 mg/mL (in PBS) was injected in amount 40 μ L per well. Immediately after that, 10 μ L of given concentration of reactivator (or PBS) was injected per well and the absorbance shift at 412 nm was followed using MRX device (Dynatech Laboratories, Chantilly, VA, USA). Absorbance was measured after 15 min. Obtained experimental data were processed according to the following equation:

$$R = \frac{A_r - A_b}{A_0 - A_b} \times 100(\%) \quad (1)$$

The percent of reactivation (R) is expressed as function of absorbances measured in wells with reactivated BChE (A_r), intact BChE (A_0). Absorbances A_r and A_0 are compared to the blank (reaction mixture: 1 mM ATChCl with DTBN 0.4 mg/mL in PBS) absorbance - A_b .

Results and discussion

Experiment was designed as an *in vitro* study for reactivation of BChE inhibited by trichlorfon using photometric microplate assay using previously proposed protocol [18]. Reactivation *in vitro* does not typically provide a complex data since several effects in body such as partial metabolizing could influence achieved results. The presented study is targeted as *in vitro* study because of above mentioned trichlorfon metabolizing so followed reactivation of cholinesterases in tissues or organisms is typically concerning to 2,2-dimethyl dichlorovinyl phosphate when *in vivo* experiment is chosen. Our study is based only on reactivation of trichlorfon inhibited BChE because BChE is commonly useful in way of scavenger. Furthermore, it plays a deputy role in organism when Alzheimer's disease is treated by trichlorfon [23].

Inhibition of BChE was realized using four potentially effective reactivators: HI-6, obidoxime and pralidoxime are reactivators that were used in some armies as therapeutic for individuals poisoned by nerve agents; K048 is previously synthesized reactivator with properties similar to the above mentioned ones. The achieved data are digestedly summarized in Figure 2 for reactivators application within half an hour after BChE inhibition and in Figure 3 for reactivators application within one hour.

One common effect was obvious – the concentration of reactivator should reach 1 mM for effective reactivation (above 50%). Only low reactivation was observed for reactivators in concentration 0.1 mM and 0.01 mM. Efficacy of reactivation strongly decreases (R decrease about 10% for all of reactivators in concentration 1 mM) when time interval of trichlorfon and BChE preincubation is prolonged from half an hour to one hour. These facts respond to expectancies

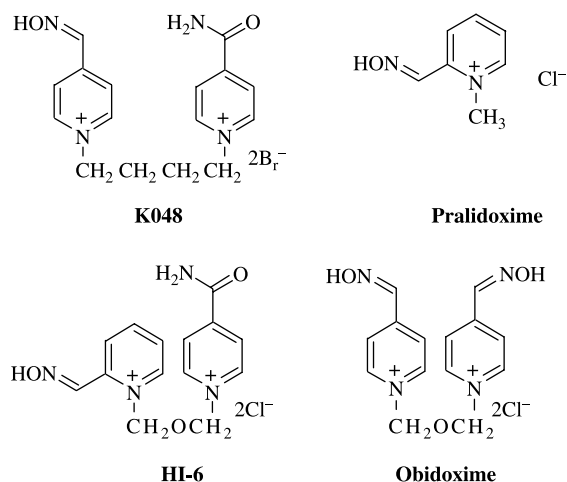


Figure 1. Reactivators used.

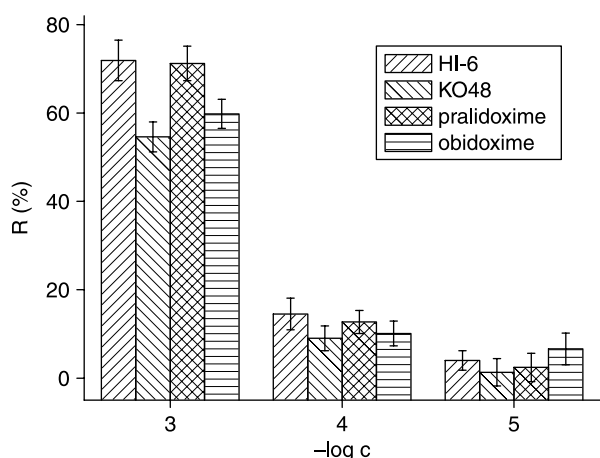


Figure 2. Semilogarithmic expression of reactivation efficacy. Percent of reactivation (R) vs. logarithm of reactivator molar concentration (final on plate). Reactivation was realized after 0.5 h incubation of trichlorfon with butyrylcholinesterase. Error bars indicate standard deviation ($n = 4$).

especially when we considered spontaneous dealkylation (aging) of trichlorfon bound in enzyme active centre and no chance to reactive dealkylated organophosphate from this centre [24,25]. The difference between half an hour and one hour preincubation before reactivation application is significant on probability level approx. 0.2.

The highest efficacy of trichlorfon inhibited BChE reactivation proved two reactivators: HI-6 and pralidoxime in concentration 1 mM. The high reactivation by HI-6 was quite surprising when we consider strong competitive effect between this reactivator and substrate not only for AChE but also for BChE [26]. We should consider reactivator concentration 1 mM as the lower limit since all reactivators in concentration 0.1 mM and lower

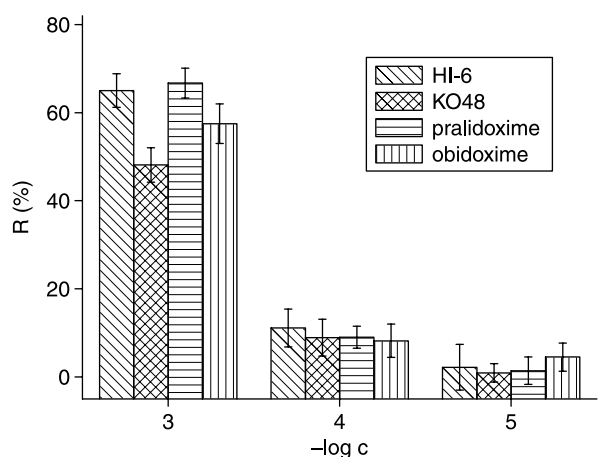


Figure 3. Semilogarithmic expression of reactivation efficacy. The meaning of symbols is the same as in Figure 2. Data were obtained for reactivation after 1 h of incubation of butyrylcholinesterase with trichlorfon.

provided efficacy under 20%. The increase of reactivator concentration strongly above 1 mM is not convenient due to starting of intoxication symptoms resulting in cholinergic crisis similar to that when individuals intoxicated by organophosphate.

An intriguing conclusion could be realized if the efficacy (percent of reactivation R) of HI-6 and pralidoxime is compared with obidoxime and KO48. Though pralidoxime and HI-6 are monoquaternary reactivators, both are more effective for reactivation purposes than bisquaternary obidoxime and monoquaternary KO48. It seems that more important for reactivation of BChE inhibited by trichlorfon purposes is position of oxime group (HI-6 and pralidoxime have oxime in position ortho in comparison with position para for obidoxime and KO48) rather than monoquaternary or bisquaternary structure.

Conclusion

The efficacy of trichlorfon-inhibited BChE treatment was followed in this study using a photometric microplate assay. The *in vitro* model was chosen to avoid the partial metabolism of trichlorfon in the body or tissues. So that the reactivation efficacy is selective for only trichlorfon.

Acknowledgements

This work was supported by the Ministry of Defence of the Czech Republic, Grant No. FVZ0000604.

Declaration of interest: The authors report no conflicts of interest. The authors alone are responsible for the content and writing of the paper.

References

- [1] Taylor P. Development of acetylcholinesterase inhibitors in the therapy of Alzheimer's disease. *Neurology* 1998;51:30–35.
- [2] McKeth IG. The clinical trial protocol of the metrifonate in Alzheimer's trial (MALT). *Dement Geriatr Cogn Disord* 1998;9:2–7.
- [3] Cummings JL, Ringman JM. Metrifonate (Trichlorfon): A review of the pharmacology, pharmacokinetics and clinical experience with a new acetylcholinesterase inhibitor for Alzheimer's disease. *Expert Opin Inv Drug* 1999;8:463–471.
- [4] Morris JC, Cyrus PA, Orazem J, Mas J, Bieber F, Ruzicka BB, Gulanski B. Metrifonate benefits cognitive, behavioral, and global function in patients with Alzheimer's disease. *Neurology* 1998;50:1222–1230.
- [5] Sun FY, Schmid TE, Schmid E, Baumgartner A, Adler ID. Trichlorfon induces spindle disturbances in V79 cells and aneuploidy in male mouse germ cells. *Mutagenesis* 2000;15:17–24.
- [6] Jann MW. Preclinical pharmacology of metrifonate. *Pharmacotherapy* 1998;18:55–67.
- [7] Pettigrew LC, Bieber F, Lettieri J, Wermeling DP, Schitt FA, Tikhtman AJ, Ashford JW, Smith CD, Wekstein DR, Markesbery WR, Orazem J, Ruzicka BB, Mas J, Gulanski B. Pharmacokinetics, pharmacodynamics, and safety of metrifonate

- in patients with Alzheimer's disease. *J Clin Pharmacol* 1998;38:236–245.
- [8] Kuca K, Jun D, Musilek K. Structural requirements of acetylcholinesterase reactivators. *Mini Rev Med Chem* 2006;6:269–277.
- [9] Wiesner J, Kriz Z, Kuca K, Jun D, Koca J. Acetylcholinesterase – the structural similarities and differences. *J Enz Inhib Med Chem* 2007;22:417–424.
- [10] Kuca K, Kassa J. A comparison of the ability of a new bispyridinium oxime-1-(4-hydroxyiminomethylpyridinium)-4-(4-carbamoylpyridinium)butane dibromide and currently used oximes to reactivate nerve agent-inhibited rat brain acetylcholinesterase by *in vitro* methods. *J Enz Inhib Med Chem* 2003;18:529–535.
- [11] Kuca K, Patocka J. Reactivation of cyclosarin-inhibited rat brain acetylcholinesterase by pyridinium-oximes. *J Enz Inhib Med Chem* 2004;19:39–43.
- [12] Kuca K, Cabal J, Kassa J. *In vitro* reactivation of sarin-inhibited brain acetylcholinesterase from different species by various oximes. *J Enz Inhib Med Chem* 2005;20:227–232.
- [13] Picha J, Kuca K, Kivala M, Kohout M, Cabal J, Liska F. A new group of monoquaternary reactivators of acetylcholinesterase inhibited by nerve agents. *J Enz Inhib Med Chem* 2005;20:233–237.
- [14] Kuca K, Cabal J, Jun D, Bajgar J, Hrabanova M. Potency of new structurally different oximes to reactivate cyclosarin-inhibited human brain acetylcholinesterase. *J Enz Inhib Med Chem* 2006;21:663–666.
- [15] Musilek K, Holas O, Kuca K, Jun D, Dohnal V, Dolezal M. Synthesis of a novel series of non-symmetrical bispyridinium compounds bearing a xylene linker and evaluation of their reactivation activity against tabun and paraoxon-inhibited acetylcholinesterase. *J Enz Inhib Med Chem* 2007;23:70–76.
- [16] Kassa J, Jun D, Kuca K. A comparison of reactivating efficacy of newly developed oximes (K074, K075) and currently available oximes (obidoxime, HI-6) in cyclosarin-and tabun-poisoned rats. *Enz Inhib Med Chem* 2007;22:297–300.
- [17] Patocka J, Kuca K, Jun D. Acetylcholinesterase and butyrylcholinesterase – important enzymes of human body. *Acta Med (Hradec Kralove)* 2004;47:215–228.
- [18] Pohanka M, Jun D, Kuca K. Photometric microplate assay for estimation of the efficacy of paraoxon-inhibited acetylcholinesterase reactivation. *J Enz Inhib Med Chem* 2007; In press.
- [19] Musilek K, Lipka L, Racakova V, Kuca K, Jun D, Dohnal V, Dolezal M. New methods in synthesis of acetylcholinesterase reactivators and evaluation of their potency to reactivate cyclosarin-inhibited. *AChE Chem Pap* 2006;60:48–51.
- [20] Musilek K, Kuca K, Jun D, Dolezal V. Synthesis of bispyridinium compounds bearing propane linker and evaluation of their reactivation activity against tabun – and paraoxon inhibited acetylcholinesterase. *Lett Org Chem* 2006;3:831–835.
- [21] Kuca K, Musilek K, Stodulka P, Marek J, Hanusova P, Jun D, Hrabanova M, Kassa J, Dolezal B. Twelve different HI-6 salts and their potency to reactivate cyclosarin inhibited AChE *in vitro*. *Lett Drug Des Discov* 2007;4:510–512.
- [22] Kuca K, Bielavsky J, Cabal J, Kassa J. Synthesis of a new reactivator of tabun inhibited acetylcholinesterase. *Bioorg Med Chem Lett* 2003;13:3545–3547.
- [23] Greig NH, Lahiri DK, Sambamurti K. Butyrylcholinesterase: An important new target in Alzheimer's disease therapy. *Int Psychogeriatr* 2002;14:77–91.
- [24] Shafferman A, Ordentlich A, Barak D, Stein D, Ariel N, Velan B. Aging of phosphorylated human acetylcholinesterase: Catalytic processes mediated by aromatic and residues of the active centre. *Biochem J* 1996;318:833–840.
- [25] Masson P, Fortier PL, Albaret C, Froment MT, Barttels CF, Lockridge O. Aging of di-isopropyl-phosphorylated human butyrylcholinesterase. *Biochem J* 1997;327:601–607.
- [26] Pohanka M, Jun D, Kuca K. Amperometric biosensor for evaluation of competitive cholinesterase inhibition by the reactivator HI-6. *Anal Lett* 2007;40:2351–2359.